

Sixty-fourth Supplement to the American Ornithological Society's *Check-list of North American Birds*

R. Terry Chesser,^{1,2,13,14} Shawn M. Billerman,³ Kevin J. Burns,⁴ Carla Cicero,⁵ Jon L. Dunn,⁶ Blanca E. Hernández-Baños,⁷ Rosa Alicia Jiménez,⁸ Andrew W. Kratter,⁹ Nicholas A. Mason,¹⁰ Pamela C. Rasmussen,^{3,11} J. V. Remsen, Jr.,¹⁰ and Kevin Winker¹²

¹U.S. Geological Survey, Eastern Ecological Science Center, 12100 Beech Forest Road, Laurel, Maryland 20708, USA;

²National Museum of Natural History, MRC-116, Smithsonian Institution, P. O. Box 37012, Washington, DC 20013-7012, USA;

³Cornell Laboratory of Ornithology, 159 Sapsucker Woods Rd., Ithaca, New York 14850, USA;

⁴Department of Biology, San Diego State University, San Diego, California 92182-4614, USA;

⁵Museum of Vertebrate Zoology, 3101 Valley Life Sciences Building, University of California, Berkeley, California 94720-3160, USA;

⁶24 Idaho Street, Bishop, California 93514, USA;

⁷Departamento de Biología Evolutiva, Facultad de Ciencias, Museo de Zoología, Universidad Nacional Autónoma de México, Apartado Postal 70-399, Ciudad de México, México;

⁸Escuela de Biología, Universidad de San Carlos de Guatemala, Ciudad de Guatemala, Guatemala;

⁹Florida Museum of Natural History, P. O. Box 117800, University of Florida, Gainesville, Florida 32611, USA;

¹⁰Museum of Natural Science and Dept. Biological Sciences, Louisiana State University, Foster Hall 119, Baton Rouge, Louisiana 70803, USA;

¹¹Michigan State University Museum and Department of Integrative Biology, West Circle Drive, East Lansing, Michigan 48824-1045, USA;

¹²University of Alaska Museum, 907 Yukon Drive, Fairbanks, Alaska 99775-6960, USA.

¹³Authors are members of the Committee on Classification and Nomenclature—North and Middle America, of the American Ornithological Society (formerly American Ornithologists' Union), listed alphabetically after the Chairman.

¹⁴E-mail: chessert@si.edu

This is the 23rd supplement since publication of the 7th edition of the *Check-list of North American Birds* (American Ornithologists' Union [AOU] 1998). It summarizes decisions made between 15 April 2022 and 25 April 2023 by the American Ornithological Society's (formerly American Ornithologists' Union) Committee on Classification and Nomenclature—North and Middle America. The Committee has continued to operate in the manner outlined in the 42nd Supplement (Banks et al. 2000). During the past year, Douglas F. Stotz left the committee.

Changes in this supplement include the following: (1) 3 species (*Psittacara erythrogenys*, *Monticola saxatilis*, and *Sporophila bouvionides*) are added to the main list on the basis of new distributional information, including one transferred from the Appendix; (2) 7 species (*Antrastomus ekmani*, *Accipiter atricapillus*, *Corvus minutus*, *Delichon lagopodum*, *Chlorophonia sclateri*, *Chlorophonia flavifrons*, and *Melopyrrha taylori*) are added to the main list because of splits from species already on the list; (3) 1 species name is changed (to *Lepidothrix velutina*) because of a split from an extralimital species; (4) the distributional statements of 3 species (*Sclerurus mexicanus*, *Chlorothraupis carmioli*, and *Amaurospiza concolor*) are changed because of splits from extralimital species; (5) 1 species (*Empidonax occidentalis*) is lost by merger with a species already on the list; (6) 1 species (*Myiodynastes chrysocephalus*) is removed due to transfer of a subspecies, rendering the species extralimital; (7) 1

genus (*Microspizias*) is added due to a split from another genus, resulting in a change to 1 scientific name (*Microspizias superciliosus*); (8) 3 genera (*Mustelirallus*, *Rufirallus*, and *Periporphyrus*) are added by merger with other genera, resulting in the loss of 4 genera (*Neocrex*, *Cyanolimnas*, *Micropygia*, and *Rhodothraupis*) and changes to 5 scientific names (*Mustelirallus colombianus*, *M. erythrops*, *M. cerverai*, *Rufirallus schomburgkii*, and *Periporphyrus celaeno*); (9) 1 genus (*Myrmothera*) is added due to a transfer of a species between genera, resulting in 1 change to a scientific name (*Myrmothera dives*); (10) a subfamily name is changed (to Florisuginae) because of nomenclatural priority; and (11) 2 species (*Falco ruficularis* and *Amazona finschi*) are added to the list of species known to occur in the United States.

New linear sequences are adopted for species in a portion of the Rallidae, for species in the genus *Psittacara*, for genera and species in the Grallariidae, and for species in a portion of the Cardinalidae, all due to new phylogenetic data.

Literature that provides the basis for the Committee's decisions is cited at the end of this supplement, and citations not already in the Literature Cited of the 7th edition (with supplements) become additions to it. A list of the bird species known from the AOS *Check-list* area can be found at <https://checklist.americanornithology.org/taxa>, and proposals that form the basis for this supplement can be found at <https://checklist.americanornithology.org/about/committees/nacc/current-prior-proposals/2023-proposals/>

The following changes to the 7th edition (page numbers refer thereto) and its supplements result from the Committee's actions:

pp. xvii–liv. Increase the number in the title of the list of species to 2,186. Insert the following names in the proper position as indicated by the text of this supplement:

Florisuginae

Antrostomus cubanensis Cuban Nightjar.
Antrostomus ekmani Hispaniolan Nightjar.
Mustelirallus cerverai Zapata Rail.
Mustelirallus colombianus Colombian Crake.
Mustelirallus erythrops Paint-billed Crake.
Rufirallus schomburgkii Ocellated Crake.
Microspizias superciliosus Tiny Hawk.
Accipiter gentilis Eurasian Goshawk. (A)
Accipiter atricapillus American Goshawk.
Psittacara erythrogenys Red-masked Parakeet. (I)
Lepidothrix velutina Velvety Manakin.
Empidonax difficilis Western Flycatcher.
Myrmothera dives Thicket Antpitta.
Corvus minutus Cuban Palm-Crow.
Corvus palmarum Hispaniolan Palm-Crow.
Delichon urbicum Western House-Martin. (A)
Delichon lagopodum Siberian House-Martin. (A)
Monticola saxatilis Rufous-tailed Rock-thrush. (A)
Chlorophonia musica Hispaniolan Euphonia.
Chlorophonia sclateri Puerto Rican Euphonia.
Chlorophonia flavifrons Lesser Antillean Euphonia.
Periporphyrus celaeno Crimson-collared Grosbeak.
Melopyrrha taylori Grand Cayman Bullfinch.
Sporophila bouvronides Lesson's Seedeater. (A)

Delete the following names:

Topazinae

Antrostomus cubanensis Greater Antillean Nightjar.
Neocrex colombiana Colombian Crake.
Neocrex erythrops Paint-billed Crake.
Cyanolimnas cerverai Zapata Rail.
Micropygia schomburgkii Ocellated Crake. (A)
Accipiter superciliosus Tiny Hawk.
Accipiter gentilis Northern Goshawk.
Lepidothrix coronata Blue-crowned Manakin.
Myiodynastes chrysocephalus Golden-crowned Flycatcher.
Empidonax difficilis Pacific-slope Flycatcher.
Empidonax occidentalis Cordilleran Flycatcher.
Hylopezus dives Thicket Antpitta.
Corvus palmarum Palm Crow.
Delichon urbicum Common House-Martin. (A)
Chlorophonia musica Antillean Euphonia.
Rhodothraupis celaeno Crimson-collared Grosbeak.

Adopt the following linear sequence for species in the genera *Mustelirallus* (formerly in *Neocrex* and *Cyanolimnas*, but see below) and *Pardirallus* (family Rallidae):

Pardirallus maculatus
Mustelirallus cerverai
Mustelirallus colombianus
Mustelirallus erythrops

Adopt the following linear sequence for species in the genus *Psittacara*:

Psittacara holochlorus
Psittacara brevipes
Psittacara strenuus
Psittacara mitratus
Psittacara erythrogenys
Psittacara finschi
Psittacara euops
Psittacara maugei
Psittacara chloropterus

Adopt the following linear sequence for species in the family Grallariidae:

Grallaricula flavirostris
Myrmothera dives
Hylopezus perspicillatus
Grallaria guatemalensis

Adopt the following linear sequence for species in the genera *Periporphyrus* (formerly in *Rhodothraupis*, but see below) and *Caryothraustes* (family Cardinalidae):

Periporphyrus celaeno
Caryothraustes poliogaster
Caryothraustes canadensis

Note: The entries below follow the current linear sequence as established in this and previous supplements, although entries continue to be cross-referenced to page numbers in [American Ornithologists' Union \(AOU\) \(1998\)](#).

1. [p. 286] The subfamily name Florisuginae has been shown to have priority over Topazinae, based on Bonaparte's ([Bonaparte 1853](#)) introduction of the group name Florisugeae for a tribe consisting of the two genera *Topaza* and *Florisuga* ([Dickinson and Gregory 2022](#)). Delete the heading Subfamily TOPAZINAE: Topazes, insert the heading Subfamily FLORISUGINAE: Jacobins, and add the following Notes after this heading:

Notes.—Subfamily previously known as Topazinae ([McGuire et al. 2009](#), [Chesser et al. 2012](#)), but Florisuginae has priority ([Dickinson and Rensen 2013](#), [Dickinson and Gregory 2022](#)).

2. [p. 271] *Antrostomus ekmani* is treated as a species separate from *A. cubanensis*. In the species account for *A. cubanensis*, change the English name to Cuban Nightjar, delete the *ekmani* group from the distributional statement, and change the existing Notes to: See comments under *A. ekmani*.

Insert the following new species account after the account for *A. cubanensis*:

Antrostomus ekmani Lönnberg. Hispaniolan Nightjar.

Antrostomus ekmani Lönnberg, 1929, Arkiv för Zoologi 20B, no. 6, p. 1, fig. 1. (near Jérémie, Haiti.)

Habitat.—Tropical Lowland Evergreen Forest, Pine Forest (0–1800 m).

Distribution.—*Resident* on Hispaniola.

Notes.—Formerly considered conspecific with *A. cubanensis*, but separated based on vocal differences (Garrido and Reynard 1998). Formerly placed in the genus *Caprimulgus*. See comments under *Antrastomus*.

3. [p. 135] Phylogenetic analyses of nuclear and mitochondrial DNA sequences have shown that the genera *Neocrex* and *Cyanolimnas* are very closely related to extralimital species *Mustelirallus albicollis* (Garcia-R *et al.* 2014, 2021; Kirchman *et al.* 2021; Brown *et al.* 2022). These findings result in the following changes:

After the heading and Notes for Family RALLIDAE: Rails, Gallinules, and Coots, insert the following new heading, citation, and Notes:

Genus *MUSTELIRALLUS* Bonaparte

Mustelirallus Bonaparte, 1856, Comptes Rendus Hebdomadaires des Séances de l'Académie des Sciences [Paris] 43: 599. Type, by monotypy, *Rallus albicollis* Vieillot.

Notes.—See comments under *M. erythropus* and *M. cerverai*.

Remove the headings Genus *NEOCREX* Sclater and Salvin, and Genus *CYANOLIMNAS* Barbour and Peters; move the citations for these genera into the synonymy of *Mustelirallus*; delete the existing Notes for *Neocrex*; change *Neocrex colombiana* Bangs to *Mustelirallus colombianus* (Bangs), *Neocrex erythropus* (Sclater) to *Mustelirallus erythropus* (Sclater), and *Cyanolimnas cerverai* Barbour and Peters to *Mustelirallus cerverai* (Barbour and Peters); make the appropriate changes in generic abbreviations within the existing Notes; and place the accounts for these species, in the sequence indicated below, under the heading for *Mustelirallus*. Replace the existing Notes for *M. colombianus* with the following:

Notes.—Formerly, along with *M. erythropus*, placed in the genus *Neocrex*, but genetic data indicate that *erythropus* is closely related to and should be considered congeneric with *Mustelirallus albicollis* (Vieillot, 1819) [Ash-throated Crake] (Garcia-R *et al.* 2014, 2021; Kirchman *et al.* 2021; Depino *et al.* 2023); presumably this also applies to the very similar *M. colombianus*, which has not been sampled genetically.

Insert the following Notes at the end of the species account for *M. cerverai*:

Notes.—Formerly placed in the genus *Cyanolimnas*, but genetic data indicate that *M. cerverai* is closely related to and should be considered congeneric with *Mustelirallus albicollis* (Vieillot, 1819) [Ash-throated Crake] (Brown *et al.* 2022). Similarities in phenotype (e.g., red-orange legs and base of the bill) are also apparent in the species formerly in *Cyanolimnas* and *Neocrex* and now placed in *Mustelirallus*.

4. [pp. 135–136] Phylogenetic analyses of nuclear and mitochondrial sequence data have shown that the current linear sequence of species in the genera *Mustelirallus* (formerly in *Neocrex* and *Cyanolimnas*) and *Pardirallus* does not reflect their evolutionary relationships.

Under the heading Family RALLIDAE: Rails, Gallinules, and Coots, add the following to the existing Notes: Linear sequence of *Pardirallus* and *Mustelirallus* follows Kirchman *et al.* (2021) and Brown *et al.* (2022).

Rearrange the linear sequence of species in *Pardirallus* and *Mustelirallus* to:

Pardirallus maculatus
Mustelirallus cerverai
Mustelirallus colombianus
Mustelirallus erythropus

5. [p. 129] Phylogenetic analyses of nuclear and mitochondrial DNA sequences indicate that *Micropygia schomburgkii* is the sister species to extralimital species *Rufirallus viridis*, and that these species are best placed together in *Rufirallus* (Kirchman *et al.* 2021, Depino *et al.* 2023). Replace the heading, citation, and Notes for Genus *MICROPYGIA* Bonaparte with the following:

Genus *RUFIRALLUS* Bonaparte

Rufirallus Bonaparte, 1856, Comptes Rendus Hebdomadaires des Séances de l'Académie des Sciences [Paris] 43: 599. Type, by subsequent designation (Sclater and Salvin, 1868), *Rallus cayannensis* Gmelin = *Rallus cayannensis* Linnaeus = *Rallus viridis* Müller.

Notes.—*Rufirallus schomburgkii* was formerly placed in *Micropygia*, but genetic data (Kirchman *et al.* 2021, Depino *et al.* 2023) show that this species is sister to *Rufirallus viridis* (Müller, 1776) [Russet-crowned Crake]. These species share features of plumage, soft part colors, and vocalizations, as well as a preference for drier habitats (Depino *et al.* 2023). Thus, we follow Kirchman *et al.* (2021) and Depino *et al.* (2023) in treating them as congeneric.

Change *Micropygia schomburgkii* (Schomburgk) to *Rufirallus schomburgkii* (Schomburgk), move the citation for *Micropygia* into the synonymy of *Rufirallus*, and change the distributional statement to the following:

Resident locally in southern Costa Rica (Puntarenas), southeastern Colombia, Venezuela, the Guianas, far southern Ecuador, eastern Peru, eastern Bolivia, Brazil (absent from forested Amazonia), northeastern Paraguay, and far northeastern Argentina.

Insert the following Notes at the end of the species account for *R. schomburgkii*:

Notes.—See comments under *Rufirallus*.

6. [p. 93] Analyses of osteological characters (Olson 2006) and nuclear and mitochondrial DNA sequences (Kocum 2006, Hugall and Stuart-Fox 2012, Oatley *et al.* 2015, Mindell *et al.* 2018) indicate that *Accipiter superciliosus* is not closely related to the other species in this genus, with the presumed exception of *A. collaris* (Mindell *et al.* 2018). These findings result in the following changes:

After the species account for *Harpagus bidentatus*, insert the following new heading, citation, and Notes:

Genus *MICROSPIZIAS* Sangster *et al.*

Microspizias Sangster, Kirwan, Fuchs, Dickinson, Elliott, and Gregory, 2021, *Vertebrate Zoology* 21: 421. Type, by original designation, *Falco superciliosus* Linnaeus.

Notes.—Formerly included in *Accipiter*, but osteological characters indicate that *M. superciliosus* is best placed in a separate genus (Olson 2006), and genetic data (Kocum 2006, Hugall and Stuart-Fox 2012, Oatley *et al.* 2015, Mindell *et al.* 2018) indicate that species of *Microspizias*, which apparently also includes the extralimital *M. collaris* (Sclater, 1860) [Semicollared Hawk] (Mindell *et al.* 2018, Sangster *et al.* 2021), are not closely related to *Accipiter sensu stricto*.

Change *Accipiter superciliosus* (Linnaeus) to *Microspizias superciliosus* (Linnaeus), move the account for this species to follow the heading, citation, and Notes for *Microspizias*, and replace the existing Notes with the following:

Notes.—See comments under *Microspizias*.

7. [p. 95] *Accipiter atricapillus* is treated as a species separate from *A. gentilis*. In the species account for *A. gentilis*, change the English name to Eurasian Goshawk and replace the existing habitat statement, distributional statement, and Notes with the following:

Habitat.—Coniferous, deciduous, and mixed forest, forest edge, open woodland, and now adapted to woodlots in urban areas, foraging also in cultivated regions; primarily in mountains in the southern portions of range.

Distribution.—Breeds in Eurasia from the British Isles (rare in Ireland), Scandinavia, northern Russia, and northern Russian Far East south to the Mediterranean region (including northern Morocco, where rare), Asia Minor, Iran, the Himalayas, eastern China, the Korean Peninsula, and Japan.

Winters throughout the breeding range, casually south to northern Africa, northern Indian plains, and Thailand.

Accidental [subspecies *gentilis*] in Labrador (Red Bay; 11 Nov. 1925; specimen, Univ. Mich. Mus. Zool. #62390; Van Tyne 1943) and [subspecies *albidus*] in Alaska (Shemya Island; 25–27 May 2001; photos; Schwitters 2008; and another individual 17 Sept. 2001; Gibson and Byrd 2007).

Notes.—Known in some Old World literature as the Goshawk. See comments under *A. atricapillus*.

Insert the following new species account after the account for *A. gentilis*:

Accipiter atricapillus (Wilson). American Goshawk.

Falco atricapillus Wilson, 1812, *American Ornithology* 6, p. 80, pl. 52, fig. 3. (Philadelphia, Pennsylvania.)

Habitat.—Coniferous, deciduous, and mixed forest, forest edge and open woodland, foraging also in cultivated regions; primarily in mountains in the southern portions of range.

Distribution.—[same as for the *atricapillus* group, except change “San Jacinto Mountains and [formerly?] Cuyumaca Mountains” to “San Bernardino Mountains”]

Notes.—Formerly (e.g., AOU 1957, 1983, 1998) considered conspecific with *A. gentilis*, but separated based on vocal and morphological differences (Sangster 2022) and polyphyly of mitochondrial DNA (Kunz *et al.* 2019). Treated

as a separate species from *A. gentilis* prior to AOU (1957), as American Goshawk.

8. [p. 110] A record of *Falco ruficularis* in the United States is recognized. Insert the following paragraph at the end of the distributional statement for this species:

Accidental in Texas (Santa Ana National Wildlife Refuge, Hidalgo County, 8 Dec. 2021–10 March 2022; photos; Gelernter *et al.* 2022).

9. [p. 235] After the species account for *Psittacara mitratus*, insert the following new species account:

Psittacara erythrogenys Lesson. Red-masked Parakeet.

Psittacara erythrogenys Lesson, 1844, *L'Écho du Monde Savant et l'Hermès* (part 2) 11, no. 34, column 486. (Guayaquil.)

Habitat.—Tropical Deciduous Forest, Gallery Forest. Introduced populations in North America in wooded areas and parks in urban settings.

Distribution.—Resident from northwestern Ecuador to northwestern Peru.

Introduced and established in California (notable populations in San Francisco, Los Angeles, and San Diego; Allen *et al.* 2016, Garrett 2018, Benson *et al.* 2021b). Introduced populations also present in Valencia, Spain; southern Florida (Chatfield-Taylor and Epps 2020); Hawaii (Pyle and Pyle 2017); Puerto Rico; and several cities in western Peru, including Lima and Arequipa (eBird data).

10. [pp. 234–235] Phylogenetic analyses of nuclear and mitochondrial DNA sequences have shown that our current linear sequence of species of *Psittacara* does not reflect their evolutionary relationships (Schirtzinger *et al.* 2012, Remsen *et al.* 2013). These findings result in the following changes:

After the heading Genus *PSITTACARA* Vigors, insert the following at the end of the existing Notes: Linear sequence of species follows Remsen *et al.* (2013).

Rearrange the sequence of species of *Psittacara* to:

Psittacara holochlorus
Psittacara brevipes
Psittacara strenuus
Psittacara mitratus
Psittacara erythrogenys
Psittacara finschi
Psittacara euops
Psittacara maugei
Psittacara chloropterus

11. [p. 243] Records of *Amazona finschi* in the United States (California) are recognized as representing an introduced established population. Replace the last paragraph of the distributional statement with the following:

Introduced and established in southern California (400–500+ individuals from Ventura County to San Diego County; Allen *et al.* 2016, Benson *et al.* 2021a); smaller introduced

populations also present in Florida and Texas (Pranty and Garrett 2011).

12. [p. 426] *Lepidothrix velutina* is treated as a species separate from *L. coronata*. Replace the species account for *L. coronata* with the following new account:

Lepidothrix velutina (Berlepsch). Velvety Manakin.

Pipra velutina Berlepsch, 1883, Ibis, p. 492. (Veragua [=Veraguas], Panama.)

Habitat.—Tropical Lowland Evergreen Forest, Secondary Forest (0–1350 m; Tropical and lower Subtropical zones).

Distribution.—Locally in Costa Rica (northwest to the Gulf of Nicoya on the Pacific slope, and the Sixaola region in the southeast), adjacent western Panama (western Chiriquí, Bocas del Toro, northern Veraguas), eastern Panama (both slopes from the Canal area eastward), and in South America in western and northern Colombia and northwestern Ecuador.

Notes.—Formerly (e.g., AOU 1983, 1998) considered conspecific with *L. coronata* (Spix, 1825) [Blue-capped Manakin], but separated based on differences in vocalizations, supported by genomic and plumage differences (Moncrieff *et al.* 2022), following Remsen *et al.* (2023).

13. [pp. 409–410] Subspecies *minor* and extralimital subspecies *cinerascens* are transferred from *Myiodynastes chrysocephalus* to *M. hemichrysus*. Remove the species account for *M. chrysocephalus*, which as newly delimited does not occur in North America. Replace the existing habitat and distributional statements and Notes for *M. hemichrysus* with the following:

Habitat.—Montane Evergreen Forest, Secondary Forest (700–1850 m; upper Tropical and Subtropical zones).

Distribution.—*Resident* on both slopes in the highlands of Costa Rica (from Cordillera de Guanacaste southward, more common on Caribbean slope), western Panama (east to Veraguas), and extreme eastern Panama (cerros Pirre, Tacarcuna, and Malí in eastern Darién), and in South America from Colombia and northern Venezuela south to Ecuador and northwestern Peru north of the Marañón Valley.

Notes.—Includes subspecies *minor* and *cinerascens*, formerly (e.g., AOU 1983, 1998) considered part of *M. chrysocephalus* (Cabanis, 1862) [Golden-bellied Flycatcher] but transferred to *M. hemichrysus* based on vocal similarities (Boesman 2016a).

14. [pp. 397–398] *Empidonax occidentalis* is treated as conspecific with *E. difficilis*. Remove the species account for *E. occidentalis*. In the species account for *E. difficilis*, change the English name to Western Flycatcher and change the habitat statement to the following:

Habitat.—Breeds in humid coniferous forest, pine-oak forest, and dense second-growth woodland (0–3500 m; Subtropical and Temperate Zones); winters in Pine-oak Forest, Montane Evergreen Forest, Gallery Forest, Deciduous Forest, Tropical Deciduous Forest, Tropical Lowland Evergreen Forest, and Tropical Lowland Evergreen Forest Edge.

In the distributional statement for *E. difficilis*, insert the *Breeds* and *Winters* distributional information from the spe-

cies account for *E. occidentalis* under [*occidentalis* group], following the distributional information for the *difficilis* group and preceding that for the *insulicola* group; delete the *Migrates* paragraph. Replace the last paragraph of the distributional statement with the following:

Casual to accidental [*difficilis* group] along the eastern seaboard from Nova Scotia (Sable Island) south to north-central Florida. Accidental [*occidentalis* group] in southern Louisiana; reports from southeastern Texas are insufficiently documented. Numerous other records in eastern North America have not been satisfactorily identified to group.

Notes.—Groups: *E. difficilis* Baird, 1858 [Pacific-slope Flycatcher], *E. occidentalis* [Cordilleran Flycatcher], and *E. insulicola* Oberholser, 1897 [Channel Islands Flycatcher]. *Empidonax difficilis* (including *insulicola*) and *E. occidentalis* were formerly (e.g., AOU 1998) considered separate species, although they had previously (e.g., AOU 1957, 1983) been considered conspecific (as Western Flycatcher). The two are again treated as a single species based on extensive hybridization and lack of consistent vocal, genomic, or morphological differentiation in a broad contact zone in southern British Columbia, southwestern Alberta, eastern Washington, Idaho, and Montana (Rush *et al.* 2009, Linck *et al.* 2019, Hopping 2022).

15. [pp. 371–372] Phylogenetic analyses of nuclear and mitochondrial DNA sequences (Carneiro *et al.* 2018) have shown that *Hylopezus* as currently constituted is paraphyletic with respect to *Myrmothera* and *Grallaricula*, and that *H. dives* is not closely related to *Hylopezus sensu stricto*. This finding results in the following changes:

After the species account for *Hylopezus perspicillatus*, insert the following new heading and citation:

Genus *MYRMOTHERA* Vieillot

Myrmothera Vieillot, 1816, Analyse d'une nouvelle ornithologie élémentaire, p. 43. Type, by subsequent designation, *G. brevicauda* = *Formicarius brevicauda* Boddaert = *Myrmornis campanisona* Hermann (Sclater, Catalogue of Birds in the British Museum 15, 1890, p. 321).

Change *Hylopezus dives* (Salvin) to *Myrmothera dives* (Salvin), move the account for this species to follow the heading and citation for *Myrmothera*, and replace the existing Notes for *M. dives* with the following:

Notes.—Formerly included in *Hylopezus*, but genetic data indicate that *Hylopezus* was paraphyletic with respect to *Myrmothera* and *Grallaricula*, and that *H. dives* is not closely related to *Hylopezus sensu stricto*.

16. [pp. 371–372] Phylogenetic analyses of nuclear and mitochondrial DNA sequences (Carneiro *et al.* 2018, 2019; Harvey *et al.* 2020) have shown that our current linear sequence of the Grallariidae does not reflect their evolutionary history.

Under the heading Family GRALLARIIDAE: Antpittas, add the following to the end of the existing Notes: Linear sequence follows Carneiro *et al.* (2018, 2019) and Harvey *et al.* (2020).

Rearrange the linear sequence of species in this family to:

Grallaricula flavirostris
Myrmothera dives
Hylopezus perspicillatus
Grallaria guatimalensis

17. [p. 354] Extralimital species *Sclerurus obscurior* is treated as a species separate from *S. mexicanus*. In the species account for *S. mexicanus*, replace the habitat statement, distributional statement, and Notes with the following:

Habitat.—Montane Evergreen Forest, Tropical Lowland Evergreen Forest (0–2300 m; Subtropical and Tropical Zones).

Distribution.—*Resident* locally in Hidalgo, eastern Puebla, Veracruz, northern and southern Oaxaca, Chiapas, Guatemala, Honduras, Costa Rica, and Panama south to Cerro Tacarcuna.

Notes.—Formerly (e.g., AOU 1983, 1998) considered conspecific with *S. obscurior* Hartert, 1901 [South American Leaf-tosser], but separated based on differences in vocalizations (Cooper and Cuervo 2017) and paraphyly of *S. mexicanus sensu lato* with respect to *S. ruficularis* Pelzeln, 1868 [Short-billed Leaf-tosser] (d'Horta *et al.* 2013, Harvey *et al.* 2020), following Remsen *et al.* (2023).

18. [p. 450] *Corvus minutus* is treated as a species separate from *C. palmarum*. In the species account for *C. palmarum*, change the English name to Hispaniolan Palm-Crow and replace the habitat and distributional statements and the existing Notes with the following:

Habitat.—Pine Forest, Tropical Lowland Evergreen Forest, Tropical Deciduous Forest (0–2000 m).

Distribution.—*Resident* on Hispaniola (mostly in the mountains).

Notes.—See comments under *C. minutus*.

Insert the following new species account preceding the account for *C. palmarum*:

Corvus minutus Gundlach. Cuban Palm-Crow.

Corvus minutus Gundlach, 1852, Boston Journal of Natural History 6(3): 315. (Cuba.)

Habitat.—Pine Forest, Palm Forest (0–600 m).

Distribution.—*Resident* on Cuba (Cienfuegos Province and locally in Camagüey Province, formerly more widespread).

Notes.—Formerly (e.g., AOU 1983, 1998) considered conspecific with *C. palmarum*, but separated based on vocal differences (Garrido *et al.* 1997, Boesman 2016b) commensurate with those between other species of *Corvus*, as well as apparent differences in display behavior (Wetmore and Swales 1931, Holyoak 1983, Raffaele *et al.* 2020).

19. [p. 463] *Delichon lagopodum* is treated as a species separate from *D. urbicum*. In the species account for *D. urbicum*, change the English name to Western House-Martin (following Leader *et al.* 2021) and change the distributional statement and Notes to the following:

Distribution.—*Breeds* widely throughout Europe including Mediterranean islands and northern Africa, through Central Asia and the Middle East north of the Arabian Peninsula, across Russia to the Pechora Region and sporadically east to about the Yenisei, through northwestern China and Mongolia, and in the southeast to northern Afghanistan, northern Kashmir, Ladakh, and Himachal Pradesh (western Himalayas of India). Casual breeder in Iceland, Namibia, and South Africa.

Winters in Mediterranean region, Cape Verde Islands (scarce), sub-Saharan Africa (mainly southeastern Africa), and less often Peninsular (mainly western) India.

Migrates between wintering and breeding grounds via Canary and Cape Verde islands, Sahara Desert, Middle East, and as far east as sub-Himalayan India.

Casual or accidental to St. Pierre et Miquelon, Greenland, Bermuda, Guadeloupe, Barbados, the Azores, Madeira, Principe, Ascension Island, St. Helena, Chagos, Lakshadweep, Maldives, Seychelles, Comoros, and northern Thailand.

Notes.—See comments under *D. lagopodum*.

Insert the following new species account after the account for *D. urbicum*:

Delichon lagopodum (Pallas). Siberian House-Martin.

Hirundo lagopoda Pallas, 1811, Zoographia Rosso-Asiatica, 1, p. 532. (Dauria.)

Habitat.—Human settlements, farmland, towns, nesting in colonies on buildings, cliffs, and bridges.

Distribution.—*Breeds* in central to northeastern Russia from just west of the Yenisei northeast to western Chukotka, Russian Far East, and south to Krasnoyarsk and the Lake Baikal region, east to the Sea of Okhotsk, and south to northern Mongolia and northeastern China.

Winters in Southeast Asia to northern Myanmar, northern Thailand, Laos, Cambodia, and southern Vietnam.

Migrates through central and eastern China (including Hong Kong) and the Korean Peninsula.

Casual to Kazakhstan, northeastern India (Arunachal Pradesh), western Japan, Taiwan, southern Thailand, and Singapore.

Casual to northern and western Alaska, mainly the Pribilof Islands and St. Lawrence Island.

Notes.—Formerly (e.g., AOU 1983, 1998) treated as conspecific with *D. urbicum*, but separated based on sympatric breeding at several sites with no evidence of hybridization (Leader *et al.* 2021), vocal and morphological differences (Leader *et al.* 2021), and genomic results that indicate apparent paraphyly of *D. urbicum sensu lato* with respect to *D. dasyptus* (Bonaparte, 1850) [Asian House-Martin] and *D. nipalense* Moore, 1854 [Nepal House-Martin] (Brown 2019).

20. [p. 497] After the species account for *Phoenicurus phoenicurus*, insert the following new heading and citation:

Genus *MONTICOLA* Boie

Monticola Boie, 1822, Isis von Oken, col. 552. Type, by subsequent designation (G. R. Gray, 1847, Genera of Birds 1, p. 220), *Turdus saxatilis* Linnaeus.

After the heading and citation for Genus *MONTICOLA* Boie, insert the following new species account:

Monticola saxatilis (Linnaeus). Rufous-tailed Rock-thrush.

[*Turdus*] *saxatilis* (Linnaeus, 1766), *Systema Naturae* (ed. 12), p. 294. (Switzerland.)

Habitat.—Dry, stony areas including heaths, scree-slopes, lava flows, crags, and mountainous areas with cliffs, canyons, or stone walls, usually with some grass or stunted trees (mostly 1500–2700 m, locally 600–3000 m). Winters in lowland grassland and savanna with rocky outcrops, coastal scrub, and degraded, open, recently burnt, and human-disturbed areas.

Distribution.—Breeds from central Portugal and Spain east discontinuously to southern France, and through the Mediterranean region, including larger Mediterranean islands and the Middle and High Atlas Mountains of northern Morocco and Algeria, to Switzerland, west-central Austria to Moldova, the Balkans, Turkey, northern Israel, Iran, and Kazakhstan to Afghanistan, northern Baluchistan, western Pakistan, and east across Mongolia and southern Russia to Lake Baikal and northwestern China east at least to Inner Mongolia and Qinghai.

Winters mainly in East Africa from Eritrea through central Tanzania and more rarely west through the Sahel belt to Senegal. Scarce in winter on coasts of northeastern Arabian Peninsula from Kuwait to Oman. Accidental in Sri Lanka.

Migrates through northern Africa, the Red Sea region, Saudi Arabia, and the Persian Gulf states, and in fall (rarely in spring) through Pakistan and western India.

Casual (mainly in spring) to northwestern Europe. Accidental to Madeira, the Canary Islands, the Seychelles, eastern China, and Japan.

Accidental to Alaska (Utqiagvik [Barrow], 24–25 June 2021; photos; Gibson *et al.* 2023).

21. [p. 584] *Chlorophonia sclateri* and *C. flavifrons* are treated as species separate from *C. musica*. In the species account for *C. musica*, change the English name to Hispaniolan Euphonia, and replace the habitat and distributional statements and the existing Notes with the following:

Habitat.—Tropical Lowland Evergreen Forest, Tropical Deciduous Forest, Montane Evergreen Forest, Secondary Forest, Tropical Lowland Forest Edge (0–2300 m).

Distribution.—Resident on Hispaniola (including Gonâve Island).

Notes.—See comments under *C. sclateri*. Formerly placed in *Euphonia* (e.g., AOU 1983, 1998), along with *C. elegantissima* and extralimital species *C. cyanocephala* (Vieillot, 1818) [Golden-rumped Euphonia], but transferred to *Chlorophonia* based on genetic data (Imfeld *et al.* 2020) that indicate that these species form a clade with species of *Chlorophonia* rather than *Euphonia*.

Insert the following new species accounts after the account for *C. musica*:

Chlorophonia sclateri (Sclater). Puerto Rican Euphonia.

Cyanophonia sclateri Sclater, 1854, *Tanagrarium Catalogus Specificus*, p. 16. (Puerto Rico.)

Habitat.—Tropical Lowland Evergreen Forest, Tropical Deciduous Forest, Secondary Forest, Tropical Lowland Evergreen Forest Edge (0–1300 m).

Distribution.—Resident on Puerto Rico.

Notes.—Formerly considered conspecific with both *C. musica* and *C. flavifrons*, but these three species separated based on striking differences in plumage (Cory 1889, Ridgway 1902, Greeney 2021) commensurate with or exceeding those found between most species pairs of euphonias.

Chlorophonia flavifrons (Sparrman). Lesser Antillean Euphonia.

Emberiza flavifrons Sparrman, 1789, *Museum Carlsonianum*, fasc. 4, no. 92, pl. 92. (St. Bartholomew, cf. Sundevall, 1869, *Öfversigt af Kongl. Vetenskaps-Akademiens Förhandlingar* 26: 583.)

Habitat.—Tropical Lowland Evergreen Forest, Tropical Deciduous Forest, Montane Evergreen Forest, Secondary Forest, Tropical Lowland Forest Edge (0–1450 m).

Distribution.—Resident in the Lesser Antilles (Antigua, Montserrat, Guadeloupe, Dominica, Martinique, St. Lucia, St. Vincent, and Grenada).

Accidental or casual (or perhaps formerly bred but now extirpated) elsewhere in the Lesser Antilles (Anguilla, St. Barthélemy, Saba, Barbuda, St. Kitts, Les Saintes, La Désirade, Bequia).

Notes.—See comments under *C. sclateri*.

22. [p. 573] Extralimital species *Chlorothraupis frenata* is treated as a species separate from *C. carmioli*. In the species account for *C. carmioli*, remove the *frenata* group from the distributional statement and replace the existing Notes with the following:

Notes.—Formerly considered conspecific with extralimital species *C. frenata* Berlepsch, 1907 [Yellow-lored Tanager], but separated based on differences in plumage and voice commensurate with differences between species pairs in related lineages (Ridgely and Greenfield 2001, del Hoyo and Collar 2016, Hilty 2022), as well as paraphyly of *C. carmioli sensu lato* with respect to *C. olivacea* (Scott 2022).

23. [pp. 632–633] Phylogenetic analyses of nuclear and mitochondrial DNA sequences (Barker *et al.* 2015, Bocalini *et al.* 2021) have shown that our current generic classification and linear placement of *Rhodothraupis celaeno* does not reflect its evolutionary relationships and that this species is closely related to extralimital species *Periporphyrus erythromelas*. This finding results in the following changes:

After the species account for *Chlorothraupis olivacea*, insert the following new heading, citation, and Notes:

Genus *PERIPORPHYRUS* Reichenbach

Periporphyrus Reichenbach, 1850, *Avium Systema Naturale*, pl. 77. Type, by subsequent designation (G. R. Gray, 1855, *Catalogue of the Genera and Subgenera of Birds Contained in the British Museum*, p. 75), *Loxia erythromelas* Gmelin.

Notes.—See comments under *P. celaeno*.

Remove the heading Genus *RHODOTHRAUPIS* Ridgway, move the citation for this genus into the synonymy of *Periporphyrus*, and delete the existing Notes for this genus. Change *Rhodothraupis celaeno* (Deppe) to *Periporphyrus celaeno* (Deppe), place the account for this species under the heading for *Periporphyrus*, and insert the following Notes at the end of the species account:

Notes.—Formerly placed in *Rhodothraupis*, but genomic data (Bocalini *et al.* 2021) indicate that *Periporphyrus* and *Rhodothraupis* are very closely related and that *Rhodothraupis* should be subsumed into *Periporphyrus*; these species also share dichromatic plumage, unlike their sister lineage *Caryothraustes*.

24. [p. 594] Extralimital species *Amaurospiza aequatorialis* is treated as a species separate from *A. concolor*. In the species account for *A. concolor*, delete southwestern Colombia and northwestern Ecuador from the distributional statement for the *concolor* group, and insert the following at the end of the existing Notes: Formerly considered conspecific with *A. aequatorialis* Sharpe, 1888 [Ecuadorian Seed-eater], but separated based on paraphyly of *A. concolor sensu lato* with respect to *A. moesta* and *A. carrizalensis*, as well as vocal and plumage differences commensurate with those between some species pairs in related lineages (Areta *et al.* 2023).

25. [p. 594] *Melopyrrha taylori* is treated as a species separate from *M. nigra*. In the species account for *M. nigra*, change the distributional statement to “Resident on Cuba (including some coastal cays) and the Isle of Pines.” and insert the following Notes at the end of the species account:

Notes.—See comments under *M. taylori*.

After the account for *Melopyrrha nigra*, insert the following new species account:

Melopyrrha taylori Hartert. Grand Cayman Bullfinch.

Melopyrrha taylori Hartert, 1896, Novitates Zoologicae 3: 257. (Grand Cayman.)

Habitat.—Tropical Deciduous Forest, Secondary Forest, Second Growth Scrub, Pastures/Agricultural Lands (0–20 m).

Distribution.—Resident on Grand Cayman Island, where widespread but rare in the west.

Accidental to Little Cayman Island (Snipe Point, 29 Sept.–18 Oct. 1998; Bradley 2000), presumably this species rather than *M. nigra*.

Notes.—Formerly (e.g., AOU 1983, 1998) considered conspecific with *M. nigra*, but separated based on differences in size, female plumage, and song (Hartert 1896, Ridgway 1901, Garrido *et al.* 2014) commensurate with or exceeding those of several related taxa of Caribbean and Galapagos finches.

26. [p. 592] Before the species account for *Sporophila lineola*, insert the following new species account:

Sporophila bouvronides (Lesson). Lesson’s Seed-eater.

Pyrrhula bouvronides Lesson, 1831, Traité d’Ornithologie, livr. 6, p. 450. (Trinidad, designated by Hellmayr 1938.)

Habitat.—Low Seasonally Wet Grasslands, Second-Growth Scrub, Riparian Thickets (0–1000 m).

Distribution.—Resident and Breeds from northeastern Colombia through northern Venezuela, Trinidad & Tobago, Guyana, Suriname, and French Guiana. In non-breeding season occurs throughout northern and western Amazonia south and west to eastern Ecuador, eastern Peru, and northernmost Bolivia (Pando).

Accidental in Costa Rica (Pococí, Limón province, 25 June 2022; photos; Gatgens-García *et al.* 2022); several sight reports from Panama. A record from Quebec (Pointe-aux-Outardes; 24 Oct.–3 Nov. 2021) is of questionable origin (Comité d’homologation des oiseaux rares du Québec 2022).

27. [p. 697] Delete the account for *Sporophila bouvronides* from the Appendix, Part 1.

28. [pp. 705 ff.] Make the following changes to the list of French names of North American birds:

Insert the following names in the proper position as indicated by the text of this and previous supplements:

Antrostomus cubanensis Engoulevent de Cuba
Antrostomus ekmani Engoulevent d’Hispaniola
Mustelirallus cerverai Râle de Zapata
Mustelirallus colombianus Râle de Colombie
Mustelirallus erythrops Râle à bec peint
Rufirallus schomburgkii Râle ocellé
Microspizias superciliosus Épervier nain
Accipiter atricapillus Autour d’Amérique
Psittacara erythrogenys Conure à tête rouge
Lepidothrix velutina Manakin velouté
Empidonax difficilis Moucherolle obscur
Myrmothera dives Grallaire buissonnière
Corvus minutus Corneille minute
Delichon lagopodium Hirondelle de Pallas
Monticola saxatilis Monticole de roche
Chlorophonia sclateri Organiste de Porto Rico
Chlorophonia flavifrons Organiste des Petites Antilles
Periporphyrus celaeno Cardinal à collier
Tiaris olivaceus Tiaris grand-chanteur
Melopyrrha portoricensis Pèrenoir de Porto Rico
Melopyrrha grandis Pèrenoir de Saint-Kitts
Melopyrrha nigra Pèrenoir négrito
Melopyrrha taylori Pèrenoir de Grande Caïman
Melopyrrha violacea Pèrenoir petit-coq
Loxipasser anoxanthus Pèrenoir mantelé
Phonipara canora Petit-chanteur de Cuba
Loxigilla noctis Pèrenoir roug gorge
Loxigilla barbadensis Pèrenoir de Barbade
Sporophila bouvronides Sporophile faux-bouvron

Delete the following names:

Antrostomus cubanensis Engoulevent peut-on-voir
Neocrex colombiana Râle de Colombie
Neocrex erythrops Râle à bec peint
Cyanolimnas cerverai Râle de Zapata

Micropygia schomburgkii Rôle ocellé
Accipiter superciliosus Épervier nain
Lepidothrix coronata Manakin à tête bleue
Myiodynastes chrysocephalus Tyran à casque d'or
Empidonax difficilis Moucherolle côtier
Empidonax occidentalis Moucherolle des ravins
Hylopezus dives Grallaire buissonnière
Rhodothraupis celaeno Cardinal à collier
Tiaris olivaceus Sporophile grand-chanteur
Melopyrrha portoricensis Sporophile de Porto Rico
Melopyrrha grandis Sporophile de Saint-Kitts
Melopyrrha nigra Sporophile négrito
Melopyrrha violacea Sporophile petit-coq
Loxipasser anoxanthus Sporophile mantelé
Phonipara canora Sporophile petit-chanteur
Loxigilla noctis Sporophile rougegorge
Loxigilla barbadensis Sporophile de Barbade
 in APPENDIX (Part 1)
Sporophila bouvronides Sporophile faux-bouvron

Change the sequence of species in the families RALLIDAE, PSITTACIDAE, GRALLARIIDAE, and CARDINALIDAE as indicated by the text of this supplement.

Proposals considered but not accepted by the Committee include transfer of *Lampornis hemileucus* to *Prodosia*, revisions to the taxonomy of *Himantopus mexicanus*, separation of *Gygis microrhyncha* from *G. alba*, separation of *Melanerpes santacruzi* from *M. aurifrons*, merger of *Picoides dorsalis* with *P. tridactylus*, separation of *Colaptes aeruginosus* from *C. rubiginosus*, separation of *Eupsittula astec* from *E. nana*, revisions to the taxonomy of the *Psittacara holochlorus* complex, separation of *Amazona guatemalae* from *A. farinosa*, separation of *Amazona tresmariae* from *A. oratrix*, separation of *Pachyrhamphus uropygialis* from *P. major*, separation of *Poliocrania maculifer* from *P. exsul*, separation of *Xiphorhynchus aequatorialis* from *X. erythropygius*, separation of *Cyanocorax luxuosus* from *C. yncas*, separation of *Chlorospingus hypophaeus* from *C. flavigularis*, separation of *Melozona occipitalis* from *M. leucotis*, separation of *Pipilo socorroensis* from *P. maculatus*, separation of *Cacicus uropygialis* into two or three species, separation of *Icterus fuertesi* from *I. spurius*, separation of *Molothrus armenti* from *M. aeneus*, separation of *Geothlypis semiflava* into two or three species, separation of *Setophaga graysoni* from *S. pitiauyumi*, separation of *Basileuterus culicivorus* into as many as four species, separation of *Piranga flava* into as many as five species, separation of *Granatellus francescae* from *G. venustus*, separation of *Stelpnia versicolor* from *S. cucullata*, separation of *Ramphocelus icteronotus* from *R. flammigerus*, and separation of *Sporophila ophthalmica* from *S. corvina*. These rejected proposals and comments on them are available at <https://checklist.americanornithology.org/about/committees/nacc/current-prior-proposals/2023-proposals/>. Voting on a proposal to separate *Aphelocoma sumichrasti* from *A. woodhouseii* was postponed so that this change could be considered as part of a comprehensive proposal on the scrub-jay complex.

ACKNOWLEDGMENTS

The late Normand David, who passed away on 17 April 2023, served as the Committee's advisor for classical languages in

relation to scientific names. Michel Gosselin is the Committee's authority for French names, and Doug Stotz provides information on Neotropical species. Oscar Johnson, Max T. Kirsch, David Vander Pluym, and Nicholas Vinciguerra serve on the Early Professional Systematics Group for the committee. We thank J. I. Areta, L. Carneiro, J. C. Cooper, A. M. Cuervo, D. DeRaad, D. B. Donsker, K. Garrett, J. Gerbracht, D. D. Gibson, W. A. Hopping, S. N. G. Howell, M. J. Iliff, M. L. Isler, E. Linck, J. McCormack, A. E. Moncrieff, D. Ocampo, A. P. Peterson, T. S. Schulenberg, F. G. Stiles, and B. M. Winger for assistance, suggestions, and comments. The Committee has worked closely for the past two years with the IOU's Working Group on Avian Checklists (WGAC) to resolve discrepancies among global lists in their taxonomic treatments of North American birds. Most proposals reviewed for this supplement were part of this collaborative effort. We dedicate this supplement to our colleague Normand David, who was unfailingly generous in providing nomenclatural expertise on issues related both to the work of the Committee and to the research of individual committee members.

LITERATURE CITED

- Allen, L. W., K. L. Garrett, and M. C. Wimer (2016). *Los Angeles County Breeding Bird Atlas*. Los Angeles Audubon Society, Los Angeles, CA, USA.
- American Ornithologists' Union (AOU) (1957). *Check-list of North American Birds*, 5th ed. American Ornithologists' Union, Baltimore, MD, USA.
- American Ornithologists' Union (AOU) (1983). *Check-list of North American Birds*, 6th ed. American Ornithologists' Union, Lawrence, KS, USA.
- American Ornithologists' Union (AOU) (1998). *Check-list of North American Birds*, 7th ed. American Ornithologists' Union, Washington, D.C., USA.
- Areta, J. I., M. J. Benítez Saldívar, M. Lentino, J. Miranda, M. Ferreira, J. Klicka, and J. Pérez-Emán (2023). Phylogenetic relationships and systematics of the bamboo-specialist *Amaurospiza* blue-seedeaters. *Ibis*. doi:10.1111/ibi.13181
- Banks, R. C., C. Cicero, J. L. Dunn, A. W. Kratter, H. Ouellet, P. C. Rasmussen, J. V. Remsen, Jr., J. A. Rising, and D. F. Stotz. (2000). Forty-second supplement to the American Ornithologists' Union *Check-list of North American Birds*. *The Auk* 117:847–858.
- Barker, F. K., K. J. Burns, J. Klicka, S. M. Lanyon, and I. J. Lovette (2015). New insights into New World biogeography: An integrated view from the phylogeny of blackbirds, cardinals, sparrows, tanagers, warblers, and allies. *The Auk: Ornithological Advances* 132:333–348.
- Benson, T. A., K. L. Garrett, J. S. Feenstra, J. F. Garrett, K. N. Nelson, and A. J. Searcy. (2021a). *California Bird Records Committee proposal to add Lilac-crowned Parrot to the California state list*. California Bird Records Committee, Camarillo, CA, USA.
- Benson, T. A., K. L. Garrett, J. S. Feenstra, J. F. Garrett, K. N. Nelson, and A. J. Searcy (2021b). *California Bird Records Committee proposal to add Red-masked Parakeet to the California state list*. California Bird Record Committee, Camarillo, CA, USA.
- Bocalini, F., S. D. Bolívar-Leguizamón, L. F. Silveira, and G. A. Bravo (2021). Comparative phylogeographic and demographic analyses reveal a congruent pattern of sister relationships between bird populations of the northern and south-central Atlantic Forest. *Molecular Phylogenetics and Evolution* 154:106973.
- Boesman, P. (2016a) Notes on the vocalizations of Golden-crowned Flycatcher (*Myiodynastes chrysocephalus*) and Golden-bellied Flycatcher (*Myiodynastes hemichrysus*). HBW Alive Ornithological Note 141. In *Handbook of the Birds of the World Alive*. Lynx Edicions, Barcelona, Spain. <https://doi.org/10.2173/bow-on.100141>

- Boesman, P. (2016b) Notes on the vocalizations of Palm Crow (*Corvus palmarum*). HBW Alive Ornithological Note 209. In *Handbook of the Birds of the World Alive*. Lynx Edicions, Barcelona, Spain. <https://doi.org/10.2173/bow-on.100209>
- Bonaparte, C. L. (1853). Classification ornithologique par series. *Comptes Rendus Hebdomadaires des Séances de l'Académie des Sciences* 37(18):641–647.
- Bradley, P. (2000) *The Birds of the Cayman Islands*. BOU Checklist No. 19. British Ornithologists' Union, Tring, UK.
- Brown, C. E. (2019). *Phylogeny and evolution of swallows (Hirundinidae) with a transcriptomic perspective on seasonal migration*. Ph.D. dissertation, Department of Biological Science, Louisiana State University, Baton Rouge, LA, USA.
- Brown, A. F., Y. Lawrie, T. J. Shannon, J. M. Collinson, G. M. Kirwan, A. Kirkconnell, and M. Stervander (2022). First genetic data for the critically endangered Cuban endemic Zapata Rail *Cyanolimnas cerverai*, and the taxonomic implications. *Journal of Ornithology* 163:945–952.
- Carneiro, L., G. A. Bravo, and A. Aleixo (2019). Phenotypic similarity leads to taxonomic inconsistency: A revision of the lowland's antpittas. *Zoologica Scripta* 48:46–56.
- Carneiro, L., G. A. Bravo, N. Aristizábal, and A. Aleixo (2018). Molecular systematics and biogeography of lowland antpittas (Aves, Grallariidae): The role of vicariance and dispersal in the diversification of a widespread Neotropical lineage. *Molecular Phylogenetics and Evolution* 120:375–389.
- Chatfield-Taylor, W. E., and S. A. Epps (2020). Population trends in the exotic Red-masked Parakeet (*Psittacara erythrogenys*) in southern Florida. *Florida Field Naturalist* 48:33–44.
- Chesser, R. T., R. C. Banks, F. K. Barker, C. Cicero, J. L. Dunn, A. W. Kratter, I. J. Lovette, P. C. Rasmussen, J. V. Remsen, Jr., J. D. Rising, D. F. Stotz, and K. Winker (2012). Fifty-third supplement to the American Ornithologists' Union *Check-list of North American Birds*. *The Auk* 129:573–588.
- Comité d'homologation des oiseaux rares du Québec (2022). Rapport du comité 2022. Regroupement QuébecOiseaux, Montreal, QC, Canada. https://cdn.ca.yapla.com/company/CYPcN9TslMHKJSLwzd8PIVrDo/asset/files/CHORQ_Rapport%20du%20comite%CC%81_2022%20QO_opt.pdf
- Cooper, J. C., and A. M. Cuervo (2017). Vocal variation and species limits in the *Sclerurus mexicanus* complex. *The Wilson Journal of Ornithology* 129:13–24.
- Cory, C. B. (1889). *The Birds of the West Indies*. Estes and Lauriat, Boston, MA, USA.
- Depino, E. A., J. L. Pérez-Emán, E. Bonaccorso, and J. I. Areta (2023). Evolutionary history of New World crakes (Aves: Rallidae) with emphasis on the tribe Laterallini. *Zoologica Scripta*. <https://doi.org/10.1111/zsc.12595>.
- d'Horta, F. M., A. M. Cuervo, C. C. Ribas, R. T. Brumfield, and C. Y. Miyaki (2013). Phylogeny and comparative phylogeography of *Sclerurus* (Aves: Furnariidae) reveal constant and cryptic diversification in an old radiation of rain forest understory specialists. *Journal of Biogeography* 40:37–49.
- del Hoyo, J., and N. J. Collar (2016). *HBW and BirdLife International Illustrated Checklist of the Birds of the World. Volume 2: Passerines*. Lynx Edicions, Barcelona, Spain.
- Dickinson, E. C., and S. M. S. Gregory (2022). The correct name for a hummingbird subfamily. *Avian Systematics* 1:N7–N8.
- Dickinson, E. C., and J. V. Remsen, Jr. (Editors) (2013). *The Howard and Moore Complete Checklist of the Birds of the World, Volume 1. Non-passerines*. Aves Press, Eastbourne, UK.
- García-R, J. C., G. C. Gibb, and S. A. Trewick (2014). Deep global evolutionary radiation in birds: diversification and trait evolution in the cosmopolitan bird family Rallidae. *Molecular Phylogenetics and Evolution* 81:96–108.
- García-R, J. C., and N. J. Matzke (2021). Trait-dependent dispersal in rails (Aves: Rallidae): Historical biogeography of a cosmopolitan bird clade. *Molecular Phylogenetics and Evolution* 159:107106.
- Garrett, K. (2018). Introducing change: A current look at naturalized bird species in western North America. In *Trends and Traditions: Avifaunal Change in Western North America* (W. D. Shuford, R. E. Gill, Jr., and C. M. Handel, Editors). Studies of Western Birds 3. Western Field Ornithologists, Camarillo, CA, USA. pp. 116–130.
- Garrido, O. H., and G. B. Reynard (1998). Is the Greater Antillean Nightjar, *Caprimulgus cubanensis* (Aves: Caprimulgidae), a composite species? *Ornitología Neotropical* 9:1–12.
- Garrido, O. H., G. B. Reynard and A. Kirkconnell (1997) Is the Palm Crow, *Corvus palmarum* (Aves: Corvidae) a monotypic species? *Ornitología Neotropical* 8:15–21.
- Garrido, O. H., J. W. Wiley, P. Bradley, A. Günther-Calhoun, and D. Rodríguez (2014). Revision of the endemic West Indian genus *Melopyrrha* from Cuba and the Cayman Islands. *Bulletin of the British Ornithologists' Club* 134: 134–144.
- Gatgens-García, J., A. Chaves-Sánchez, and L. Sandoval (2022). First record of Lesson's Seedeater *Sporophila bouuronides*. *Bulletin of the British Ornithologists' Club* 142:526–528.
- Gelernter, R., M. Gustafson, Z. E. Johnson, and J. R. Ramirez-Garofalo (2022). First U.S. record of Bat Falcon. *North American Birds* 73:4–7.
- Gibson, D. D., and G. V. Byrd (2007). *Birds of the Aleutian Islands, Alaska*. Nuttall Ornithological Club and American Ornithologists' Union, Cambridge, MA, and Washington, D.C., USA.
- Gibson, D. D., S. C. Hinl, T.G. Tobish, Jr., A. J. Lang, L. H. DeCicco, N. R. Hajdukovich, and R. L. Scheer (2023). Fifth report of the Alaska Checklist Committee 2018–2022. *Western Birds* 54: in press.
- Greeney, H. (2021). Antillean Euphonia (*Chlorophonia musica*), version 2.1. In *Birds of the World* (B. K. Keeney and J. Gerbracht, Editors). Cornell Lab of Ornithology, Ithaca, NY, USA. <https://doi.org/10.2173/bow.anteup1.02.1>
- Hartert, E. (1896). Description of a new finch from the West Indies. *Novitates Zoologicae* 3:257.
- Harvey, M. G., G. A. Bravo, S. Claramunt, A. M. Cuervo, G. E. Derryberry, J. Battilana, G. F. Seeholzer, J. S. McKay, B. C. Faircloth, S. V. Edwards, J. Pérez-Emán, R. G. Moyle, F. H. Sheldon, A. Aleixo, B. T. Smith, R. T. Chesser, L. F. Silveira, J. Cracraft, R. T. Brumfield, and E. P. Derryberry (2020). The evolution of a tropical biodiversity hotspot. *Science* 370:1343–1348.
- Hilty, S. (2022). Carmiol's Tanager (*Chlorothraupis carmioli*), version 1.1. In *Birds of the World* (N. D. Sly, Editor). Cornell Lab of Ornithology, Ithaca, NY, USA. <https://doi.org/10.2173/bow.olitan1.01.1>
- Holyoak, D. T. (1983). Notes on the Palm Crow *Corvus palmarum* in Haiti. *Bulletin of the British Ornithologists' Club* 103:81–82.
- Hopping, A. (2022). Unraveling Western Flycatchers: A case against the split. *North American Birds* 73:8–17.
- Hugall, A. F., and D. Stuart-Fox (2012). Accelerated speciation in colour-polymorphic birds. *Nature* 485:631–634.
- Imfeld, T. S., F. K. Barker, and R. T. Brumfield (2020). Mitochondrial genomes and thousands of ultraconserved elements resolve the taxonomy and historical biogeography of the *Euphonia* and *Chlorophonia* finches (Passeriformes: Fringillidae). *The Auk: Ornithological Advances* 137:ukaa016.
- Kirchman J. J., N. R. McInerney, T. C. Giarla, S. L. Olson, E. Slikas, and R. C. Fleischer (2021). Phylogeny based on ultra-conserved elements clarifies the evolution of rails and allies (Ralloidea) and is the basis for a revised classification. *Ornithology* 138:ukab042.
- Kocum, A. (2006). *Phylogenie der Accipitriformes (Greifvögel) anhand verschiedener nuklearer und mitochondrialer DNA-Sequenzen*. Ph.D. Thesis, Universität Greifswald, Germany.
- Kunz, F., A. Gamauf, F. E. Zachos, and E. Haring (2019). Mitochondrial phylogenetics of the goshawk *Accipiter [gentilis]* superspecies. *Journal of Zoological Systematics and Evolutionary Research* 57:942–958.
- Leader, P., G. Carey, and M. Schweizer (2021). The identification, taxonomy and distribution of Western, Siberian and Asian House Martins. *British Birds* 114:72–96.

- Linck, E., K. Epperly, P. van Els, G. M. Spellman, R. W. Bryson, J. E. McCormack, R. Canales-del-Castillo, and J. Klicka (2019). Dense geographic and genomic sampling reveals paraphyly and a cryptic lineage in a classic sibling species complex. *Systematic Biology* 68:956–966.
- McGuire, J. A., C. C. Witt, J. V. Remsen, Jr., R. Dudley, and D. L. Altshuler (2009). A higher-level taxonomy for hummingbirds. *Journal of Ornithology* 150:155–165.
- Mindell, D. P., J. Fuchs, and J. A. Johnson (2018). Phylogeny, taxonomy, and geographic diversity of diurnal raptors: Falconiformes, Accipitriformes, and Cathartiformes. In *Birds of Prey* (J. H. Sarasola, J. Grande, and J. Negro, Editors). Springer, Cham, Switzerland.
- Moncrieff, A. E., B. C. Faircloth, and R. T. Brumfield (2022). Systematics of *Lepidothrix* manakins (Aves: Passeriformes: Pipridae) using RADcap markers. *Molecular Phylogenetics and Evolution* 173:107525.
- Oatley, G., R. E. Simmons, and J. Fuchs (2015). A molecular phylogeny of the harriers (*Circus*, Accipitridae) indicate [*sic*] the role of long distance dispersal and migration in diversification. *Molecular Phylogenetics and Evolution* 85:150–160.
- Olson, S. L. (2006). Reflections on the systematics of *Accipiter* and the genus for *Falco superciliosus* Linnaeus. *Bulletin of the British Ornithologists' Club* 126:69–70.
- Pranty, B., and K. L. Garrett (2011). Under the radar: “Non-countable” exotic species in the ABA Area. *Birding* 43:46–58.
- Pyle, R. L., and P. Pyle (2017). *The Birds of the Hawaiian Islands: Occurrence, History, Distribution, and Status, Volume 2* (tinyurl.com/Pyle-Pyle). B.P. Bishop Museum, Honolulu, HI, USA.
- Raffaele, H. A., J. W. Wiley, O. H. Garrido, A. R. Keith and J. I. Raffaele (2020). *Birds of the West Indies*, 2nd edition. Princeton Field Guides. Princeton University Press, Princeton, NJ, USA.
- Remsen, J. V., Jr., J. I. Areta, E. Bonaccorso, S. Claramunt, A. Jaramillo, D. F. Lane, J. F. Pacheco, M. B. Robbins, F. G. Stiles, and K. J. Zimmer (2023). *A classification of the bird species of South America*. American Ornithological Society, Chicago, IL, USA. <http://www.museum.lsu.edu/~Remsen/SACCBaseline.htm>
- Remsen, J. V., Jr., E. E. Schirtzinger, A. Ferraroni, L. F. Silveira, and T. F. Wright (2013). DNA-sequence data require revision of the parrot genus *Aratinga* (Aves: Psittacidae). *Zootaxa* 3641:296–300.
- Ridgely, R. S., and P. J. Greenfield (2001). *The Birds of Ecuador, Volumes 1–2*. Cornell University Press, Ithaca, NY, USA.
- Ridgway, R. (1901). *The Birds of North and Middle America, Part 1*. Bulletin of the United States National Museum No. 50, U.S. Government Printing Office, Washington, D.C., USA.
- Ridgway, R. (1902). *The Birds of North and Middle America, Part 2*. Bulletin of the United States National Museum No. 50, U.S. Government Printing Office, Washington, D.C., USA.
- Rush, A. C., R. J. Cannings, and D. E. Irwin (2009). Analysis of multilocus DNA reveals hybridization in a contact zone between *Empidonax* flycatchers. *Journal of Avian Biology* 40:614–624.
- Sangster, G. (2022). The taxonomic status of Palearctic and Nearctic populations of Northern Goshawk *Accipiter gentilis* (Aves, Accipitridae): New evidence from vocalisations. *Vertebrate Zoology* 72:445–456.
- Sangster, G., G. M. Kirwan, J. Fuchs, E. C. Dickinson, A. Elliot, and S. M. S. Gregory (2021). A new genus for the Tiny Hawk *Accipiter superciliosus* and Semicollared Hawk *A. collaris* (Aves: Accipitridae), with comments on the generic name for the Crested Goshawk *A. trivirgatus* and Sulawesi Goshawk *A. griseiceps*. *Vertebrate Zoology* 71:419–424.
- Schirtzinger, E. E., E. S. Tavares, L. A. Gonzales, J. R. Eberhard, C. Y. Miyaki, J. J. Sanchez, A. Hernandez, H. Mueller, G. R. Graves, R. C. Fleischer, and T. F. Wright (2012). Multiple independent origins of mitochondrial control region duplications in the order Psittaciformes. *Molecular Phylogenetics and Evolution* 64:342–356.
- Schwitters, M. T. (2008). *Bird species found at Shemya Island, Alaska, 1999–2007*. US Fish & Wildlife Service Report AMNWR 08/15, Homer, AK, USA.
- Scott, B. (2022). *Phylogenetics of Cardinalidae and the impact of habitat, climate, and ecology on the evolution of color*. Master's thesis, San Diego State University, San Diego, CA, USA.
- Van Tyne, J. (1943). A peculiar Goshawk from Labrador. *The Auk* 60:267–268.
- Wetmore, A., and B. H. Swales (1931). *The Birds of Haiti and the Dominican Republic*. United States National Museum Bulletin 155. U.S. Government Printing Office, Washington, D.C., USA.